BK Virus-Associated Urinary Bladder Cancer in a Kidney Transplant Recipient: A Case Report and Review of the Pathogenesis

Nemer Junior El-Mouallem, MD, and Asit K. Paul, MD, PhD

Abstract

The reactivation of polyomavirus hominis 1, more commonly known as BK virus (BKV), in solid organ transplant recipients may lead to several clinical consequences, including urinary bladder cancer. We present a case of a BKV-positive urothelial carcinoma of the urinary bladder in a 38-year-old female who received a live donor kidney transplant 15 months prior to her cancer diagnosis. The patient was treated with a radical surgery including cystectomy. Both urothelial carcinoma and carcinoma in situ portions of the surgical tissue obtained from the cystectomy specimen showed strong and diffuse expression of BKV T-Ag (tumor-antigen) by immunohistochemical staining. Although they are uncommon, reported cases of BKV-positive bladder cancer in solid organ transplant recipients have been on the rise. We review proposed pathogeneses and mechanisms of oncogenesis by which BKV can lead to tumor formation. Despite the growing body of data, the carcinogenic potential of the BK virus remains controversial.

AJHO. 2017;13(3):15-20

Introduction

Polyomavirus hominis 1, more commonly known as BK virus (BKV), infects up to 90% of the general population.¹ It is typically contracted during childhood and may remain latent in the urinary tract.² It has a subclinical or nonspecific course in immunocompetent individuals. After an asymptomatic initial infection, the virus disseminates and establishes a persistent infection in the urinary tract. With the advent of the use of immunosuppressive regimens required to maintain the viability of donor organ transplants, BKV may reactivate in the urinary tract and cause an array of clinical syndromes. Approximately one-third of renal transplant recipients on immunosuppressive medications experience urothelial BKV reactivation.³ This can lead to BKV allograft rejection, ureteral stenosis, and hemorrhagic cystitis.^{3,4} Several case reports of urothelial carcinoma have been described in renal transplant patients with

high levels of BKV reactivation.^{1,5:22} This has raised the question of a potential role for BKV in the development of urinary tract tumors in the immunosuppressed solid organ transplant population. We present a case of a BKV T-Ag (tumor-antigen) positive urothelial bladder carcinoma diagnosed in a renal transplant recipient. We also summarize a literature review of the published case reports of BKV-positive urothelial carcinomas.^{1,5:22}

Case Report

Our patient is a 38-year-old female who underwent a living donor kidney transplant in March 2014 for end-stage renal disease caused by long-standing reflux nephropathy. She had been maintained on an immunosuppressive regimen that included cyclosporine, mycophenolate mofetil, and prednisone. In June 2015, she developed painless gross hematuria, at which time she underwent a cystoscopy that revealed several bladder polyps. Transurethral resection of the bladder tumor revealed a high-grade urothelial carcinoma. Imaging with computed tomography revealed a markedly lobulated wall thickening of the antero-superior urinary bladder with a suggestion of local invasion into the abdominal wall musculature (Figure 1). Therefore, the patient underwent anterior exenteration, which included a radical cystectomy, bilateral native nephrectomy, ureterectomy, hysterectomy, bilateral salpingo-oopherectomy, left pelvic lymph-node dissection, and the creation of an ileal conduit. Pathologic staging revealed that more than 90% of the bladder was involved by high-grade urothelial carcinoma (Figure 2). Tumor extension was noted into the vagina, uterus, and adnexa. One deep pelvic lymph node was positive for involvement by tumor. A deep soft-tissue surgical margin was involved as well. Both urothelial carcinoma and carcinoma in situ portions of the surgical tissue obtained showed strong and diffuse positivity for BKV by immunohistochemical staining of T-Ag (Figure 3). Since the surgery, the patient has been under both clinical and radiological active surveillance. She has remained on an immunosuppressive regimen. To date, the patient does not have any evidence of cancer recurrence.

Discussion

There is a growing body of evidence to support a potential oncogenic role for BKV, especially in the immunocompromised host following renal transplant. This encompasses experimen-



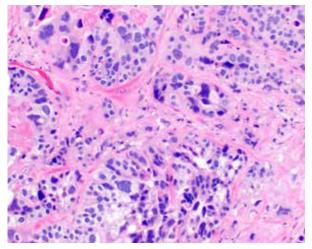
FIGURE 1: Computed Tomography Examination of the Abdomen and Pelvis.

The coronal image demonstrates the tumor (T) arising from the anterosuperior urinary bladder (UB) wall extending to the left of the midline.

tal, epidemiological and clinical evidence.¹ Once considered rare, reported cases of BKV-positive bladder cancer in solid organ transplant recipients have been on the rise (**Table 1**).

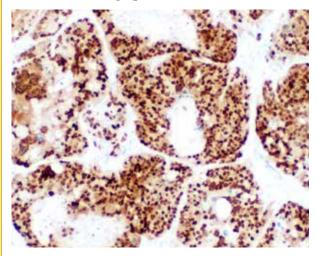
In our case, the patient developed a high-grade urothelial carcinoma that was diffusely and strongly positive for BKV. Similar to previously reported cases, the urothelial tumor developed in a solid organ transplant patient on chronic immunosuppression. Animal studies in immune-depleted or immature rodents have demonstrated the development of different tumors after a BKV challenge. The BKV T-Ag has the ability to inactivate tumor suppressor proteins of the pRB family, thereby driving the cell cycle into a proliferative state and improving the milieu for oncogenesis.²³ Moreover, the T-Ag can bind to p53, which is thought to inhibit apoptosis.²⁴ In mouse experiments, T-Ag inactivation of tumor suppressor genes pRB and p53 in the urothelium resulted in urothelial cancer resembling human carcinoma in situ.²⁵

The exact role of BKV in the development of human tumors remains a subject of debate. Although an immunocompromised state seems to enhance the oncogenic potential of BKV, bladder cancer has been observed to arise in immuFIGURE 2: High-Grade Invasive Urothelial Carcinoma Showing Nested and Sheet-Like Tumor Growth as well as Necrosis Hematoxylin-eosin stain, 200x.



More than 90% of the bladder involves high-grade urothelial carcinoma.

FIGURE 3: Immunostain Detecting Expression of BK Viral Oncoprotein (large T antigen, pAb416, Calbiochem, 1:100 dilution). Diffuse nuclear accumulation of T-antigen in viable cells (DAB chromogen, 100X) is shown in the micrograph.



Strong and diffuse positivity for BKV shown by immunohistochemical staining of T antigen.

nocompetent individuals infected with BKV as well. Among 133 immunocompetent patients diagnosed with BKV in 1 series, the most frequently occurring neoplasm was bladder cancer (15.8%).²⁵ It is still unclear whether the virus primarily represents a "passenger" or "innocent bystander" state that leads to a lytic infection, or whether it plays a direct role in

TABLE. Summary of Case Reports of BKV-Associated Urothelial Carcinomas of the Bladder Published in the Medical Literature Between 2002-2016. Superscript numbers correspond to references.

Age (years)	Gender	Type of transplant	Time From Transplant to Cancer Diagnosis	Pathology	BKV Nephropathy	Tumor cell– Specific Expression Profiles	Distinctive Features
38	F1	Cadaveric kidney- pancreas	4 years	High-grade urothelial carcinoma with glandular differentiation	Present	T-Ag, p16, p53, BerEP4, p63, CK20, β-catenin. Ki-67 83%	VP-1 DNA and mRNA detected in the tumor
60	M ¹	Kidney	11 years	High-grade urothelial carcinoma admixedwith invasive micropapillary carcinoma and adenocarcinoma in situ	Absent	T-Ag, p16,p53, BerEP4, p63, CK20, β-catenin. Ki-67 71%	VP-1 DNA was detected in tumor cells, mRNA was not
39	F⁵	Cadaveric kidney-	2 years, 2 months	Urothelial cancer	N/A	N/A	Temporal correlation was noted between high urinary BK viral loads and occurrence of bladder cancer
56	F ⁶	Cadaveric kidney	6 months	Multifocal nephrogenic adenoma of the bladder	Absent	PAX8, T-Ag, VP-1, p53. Ki-67 2%-3%	N/A
65	M ⁷	Living related- donor kidney	12 years	Pleomorphic giant cell carcinoma of the bladder (70%) admixed with areas of invasive high-grade urothelial carcinoma (20%) and trophoblastic differentiation (10%)	Absent	T-Ag, CK903, CK7, p63, p53, p16. Scattered cells positive for B-HCG	Patient had synchronous prostatic adenocarcinoma which was negative for BKV
65	F ⁸	Heart	8 years	High-grade invasive urothelial carcinoma with invasive micropapillary features	N/A	T-Ag. Ki-67>80%	VP-1 expression was negative in cancer cells, positive in inflamed urothelium
46	M ⁹	Cadaveric kidney	7 years	High-grade urothelial carcinoma	Present	T-Ag	Primary tumor and metastases stained positive for T-Ag, adjacent healthy tissue was negative
38	M ¹⁰	Cadaveric kidney	6 years	High-grade urothelial carcinoma	Present	T-Ag	Associated with diffuse carcinoma in situ
38	F ¹¹	Kidney pancreas	6.5 years	Urothelial adenocarcinoma	Present	T-Ag	N/A
55	M ¹²	Kidney- pancreas	9 years	High-grade invasive urothelial carcinoma, micropapillary variant	Present	T-Ag	No evidence of viral particles on electron microscopy
75	F ¹²	Kidney	7 years	Multifocal invasive urothelial carcinoma	Present	T-Ag	Concomitant adenocarcinoma of the prostate, negative for BKV
57	M ¹³	Kidney	5 years	Transitional-cell carcinoma of the renal pelvis	Present	T-Ag	BKV PCR present in tumor

 β -catenin, beta-catenin; β -HCG, beta-human chorionic gonadotropin; BKV, BK virus; CK, cytokeratin; N/A, not available; PCR, polymerase chain reaction; T-Ag, tumor-antigen; VP-1, viral protein-1.

Age (years)	Gender	Type of Transplant	Time From Transplant to Cancer Diagnosis	Pathology	BKV Nephropathy	Tumor–cell Specific Expression Profiles	Distinctive Features
40	F ¹⁴	Kidney	4 years	Urothelial carcinoma	Present	T-Ag on renal biopsy, not reported on bladder cancer biopsy	N/A
33	M ¹⁵	Kidney	10 years	High-grade urothelial carcinoma with micropapillary features	N/A	T-Ag	T-Ag positivity in primary tumo and metastatic focus as well as in nondysplastic urothelium
50	M ¹⁵	Kidney	10 years	High-grade urothelial carcinoma with micropapillary features	N/A	T-Ag, p16	Urothelial cancer in situ in allograft ureter positive for T-Ag
31	M ¹⁵	Kidney	18 years	High-grade urothelial carcinoma	Present	T-Ag, p16	Subsequent renal cell carcinom (T-Ag negative) was diagnosed in kidney transplant
71	M ¹⁵	Cadaveric kidney	9 years	Urothelial carcinoma	N/A	T-Ag, p16	Invasive and in situ urothelial carcinoma was positive for T-Ag. Incidental prostate cance was negative for T-Ag
57	M ¹⁶	Kidney pancreas	4 years	High-grade urothelial carcinoma	Present	T-Ag, p53	T-Ag positivity noted in both bladder tumor and metastatic focus
56	M ¹⁷	Kidney	6.5 years	High-grade urothelial carcinoma	Present	T-Ag	Primary tumor and metastases were positive for T-Ag
64	F ¹⁸	Kidney	5 years	High-grade invasive urothelial carcinoma	Present	T-Ag, p53, CK7, PAX8, p63, p16	Renal graft was obtained from pediatric donor
59	M ¹⁹	Kidney	4 years	2 separate high- grade papillary urothelial carcinomas in the renal pelvis	Present	T-Ag	BKV PCR present in the primar tumor. T-Ag was positive in both tumor and normal rena parenchyma
47	M ²⁰	Kidney	4.5 years	High-grade urothelial carcinoma in the ureter graft	Present	T-Ag	N/A
54	M ²¹	Cadaveric kidney	9 years	High-grade urothelial carcinoma in the renal pelvis	Absent	T-Ag	T-Ag positivity seen in both invasive and in situ tumor cells Negative staining for VP-1 capsid protein
65	M ²²	Heart	6 years	High-grade urothelial carcinoma with micropapillary features	N/A	T-Ag, p16, p53	T-Ag positivity in cancer cells and inflamed urothelium. VP-1 expression was positive in inflamed urothelium, negative in cancer cells

TABLE. Summary of Case Reports of BKV-Associated Urothelial Carcinomas of the Bladder Published in the Medical Literature Between 2002-2016. Superscript numbers correspond to references (continued).

β-catenin, beta-catenin; β-HCG, beta-human chorionic gonadotropin; BKV, BK virus; CK, cytokeratin; N/A, not available; PCR, polymerase chain reaction; T-Ag, tumor-antigen; VP-1, viral protein-1.

oncogenesis.^{1,22} Recent data have shown that the urotheliumspecific expression of polyomavirus SV40 T-Ag in transgenic mice produced tumors strongly resembling carcinoma in situ lesions of the human urinary bladder.¹ It has been hypothesized that the release of progeny BK virions from cell nuclei leading to cell lysis (the lytic infection phase) is considered to counteract the hypothesis of viral tumor induction and oncogenesis. However, there may be a unifying hypothesis at play: Although lytic infection and oncogenic infection are not able to co-exist at the single-cell level, cell populations can be affected by the virus differently at the whole-tissue level.

One population of cells may include the permissive urothelial cells that allow the infective lytic cycle to occur, while another cell population may include the nonpermissive cells, where "uncoupling" is the main event at play. Uncoupling refers to the virus aborting the process of cell lysis in favor of oncogenesis.²⁶

Alexiev et al¹ assembled these observations on pathogenesis into 1 unified model. In summary, this model proposes that BKV reactivates in the urinary tract in the setting of immunosuppression. This inactivates p53 and pRb, leading to G2/S (gap 2/synthesis) phase initiation. One of 2 possible sets of developments ensues thereafter. The first is a productive infection in permissive cells, leading to cell lysis, with no eventual tumor growth seen. The offshoot to this scenario is that neighboring cells can get infected as well. With time, a portion of these cells become nonpermissive, aborting the process of virion assembly and cell lysis in favor of oncogenesis. The second scenario plays out as activation of viral mutations and viral replication, which leads to a cascade of host cell genomic instability and, ultimately, neoplasia.

VP-1 is a late gene region viral protein that is the main component of the BKV capsid.^{22,27} Its expression in tumor cells may be more indicative of a lytic infection, while its lack of expression may be suggestive of an oncogenic role via the process of uncoupling. Two cases of BKV-associated urinary bladder carcinoma in renal transplant recipients reported by Alexiev et al had VP-1 DNA detected in the tumor cells by polymerase chain reaction. VP-1 messenger RNA (mRNA) was detected in 1 of these cases and was missing in the other.¹ Their data did not allow the determination of whether the viral sequences were integrated into the host genome. However, a recent case published by Kenan et al documented integration of BKV genome into cancer cells, and it presented evidence for molecular uncoupling of T-Ag expression from viral assembly via robust expression of T-Ag and prohibition of VP-1 expression.²¹ Another recent case reported by Alexiev et al²² noted T-Ag and VP-1 expression in BKV related cystitis surrounding malignant areas; however, VP-1 expression and cytopathic changes were lost in areas of malignant transformation, coupled with persistent expression of T-Ag in tumor cells.²² Lavien et al reported a BKV-associated invasive urothelial carcinoma in a cardiac transplant patient on immunosuppression. In this

instance, T-Ag expression remarkably was evident in both cancer tissue and inflamed bladder tissue; however, VP-1 expression was found only in the inflamed urothelium and not in the carcinoma tissue.⁸ These studies provide further support for the central role that the large T-Ag plays in oncogenesis.

Conclusion

Despite the rising number of case reports describing BKV-associated urothelial carcinomas and the growing body of data to support this possible viral oncogenic role, there remains a need for conclusive evidence of BKV involvement in carcinogenesis, and an elucidation of the pathophysiologic mechanisms leading to tumor development. The observed association of BKV infection with urothelial carcinoma in solid organ transplant recipients merits clinical awareness for early diagnosis and management of these malignancies.

Disclosure: This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Financial disclosures: The authors report no relationship or financial interest with any entity that would pose a conflict of interest with the subject matter of this article.

Author affiliations: Nemer Junior El-Mouallem, MD, and Asit K. Paul, MD, PhD, are with the Department of Medicine, Division of Hematology, Oncology and Palliative Care, Virginia Commonwealth University Medical Center and Massey Cancer Center, Richmond, Virginia.

Send correspondence to: Asit K. Paul, MD, PhD, Department of Medicine, Division of Hematology, Oncology and Palliative Care, Virginia Commonwealth University Medical Center, 1101 E Main St, Box 980230, Richmond, VA 23298. E-mail: asit.paul@VCUhealth.org.

References

1. Alexiev BA, Randhawa P, Vazquez Martul E, et al. BK virus-associated urinary bladder carcinoma in transplant recipients: report of 2 cases, review of the literature, and proposed pathogenetic model. *Hum Pathol.* 2013;44(5):908-917. doi: 10.1016/j.humpath.2012.09.019.

2. Chesters PM, Heritage J, McCance DJ. Persistence of DNA sequences of BK virus and JC virus in normal human tissues and in diseased tissues. *J Infect Dis.* 1983;147(4):676-684.

3. Drachenberg CB, Hirsch HH, Ramos E, Papadimitriou JC. Polyomavirus disease in renal transplantation: review of pathological findings and diagnostic methods. *Hum Pathol.* 2005;36(12):1245-1255. Review.

4. Gardner SD, Field AM, Coleman DV, Hulme B. New human papovavirus (B.K.) isolated from urine after renal transplantation. *Lancet.* 1971;1(7712):1253-1257.

5. Yin WY, Lee MC, Lai NS, Lu MC. BK virus as a potential onco-

virus for bladder cancer in a renal transplant patient. *J Formos Med Assoc.* 2015;114(4):373-374. doi: 10.1016/j.jfma.2012.02.019. 6. Alexiev BA, Papadimitriou JC, Drachenberg CB. BK polyomavirus-infected nephrogenic adenoma of the urinary bladder in a renal transplant recipient: a case report. *Pathol Res Pract.* 2015;211(9):697-701. doi: 10.1016/j.prp.2015.06.006. Review. 7. Alexiev BA, Papadimitriou JC, Chai TC, Ramos E, Staats PN, Drachenberg CB. Polyomavirus (BK)-associated pleomorphic giant cell carcinoma of the urinary bladder: a case report. *Pathol Res Pract.* 2013;209(4):255-259. doi: 10.1016/j.prp.2013.01.008. 8. Lavien G, Alger J, Preece J, Alexiev BA, Alexander RB. BK virus-associated invasive urothelial carcinoma with prominent micropapillary carcinoma component in a cardiac transplant patient: case report and review of literature. *Clin Genitourin Cancer.* 2015;13(6):e397-e399. doi: 10.1016/j.clgc.2015.04.008.

9. Hill P, Slavin J, Goodman D. High-grade urothelial carcinoma in a kidney transplant recipient with BK virus infection. *NDT Plus.* 2009;2(3):246-249. doi: 10.1093/ndtplus/sfp025.

10. Pino L, Rijo E, Nohales G, Frances A, Ubre A, Arango O. Bladder transitional cell carcinoma and BK virus in a young kidney transplant recipient. *Transpl Infect Dis.* 2013;15(1):E25-E27. doi: 10.1111/tid.12042.

11. Fernández Rivera C, Alonso Hernández A, Mosquera Reboredo J, Rodríguez Gómez I. Association of bladder adenocarcinoma and BK virus infection in a pancreatico-renal transplant recipient. *NDT Plus.* 2010;3(3):300-302.

12. Bialasiewicz S, Cho Y, Rockett R, et al. Association of micropapillary urothelial carcinoma of the bladder and BK viruria in kidney transplant recipients. *Transpl Infect Dis.* 2013;15(3):283-289. doi: 10.1111/tid.12072.

13. Saquib R, Barri YM, Saad R, et al. Transitional cell carcinoma of renal allograft associated with BK virus infection. *Dial Transpl.* 2009;38(12):518-519.

14. Roberts IS, Besarani D, Mason P, Turner G, Friend PJ, Newton R. Polyoma virus infection and urothelial carcinoma of the bladder following renal transplantation. *Br J Cancer.* 2008;99(9):1383-1386. doi: 10.1038/sj.bjc.6604711.

15. Yan L, Salama ME, Lanciault C, Matsumura L, Troxell ML. Polyomavirus large T antigen is prevalent in urothelial carcinoma post-kidney transplant. *Hum Pathol.* 2016;48:122-131. doi: 10.1016/j.humpath.2015.09.021.

16. Geetha D, Tong BC, Racusen L, Markowitz JS, Westra WH. Bladder carcinoma in a transplant recipient: evidence to implicate the BK human polyomavirus as a causal transforming agent. *Transplantation*. 2002;73(12):1933-1936.

17. van Aalderen MC, Yapici Ü, van der Pol JA, et al. Polyomavirus BK in the pathogenesis of bladder cancer. *Neth J Med.* 2013;71(1):26-28.

18. McDaid J, Farkash EA, Steele DJ, et al. Transitional cell carcinoma arising within a pediatric donor renal transplant in association with BK nephropathy. *Transplantation*. 2013;95(5):e28-e30. doi: 10.1097/TP.0b013e31828235ec.

19. Li JY, Fang D, Yong TY, Klebe S, Juneja R, Gleadle JM. Transitional cell carcinoma in a renal allograft with BK nephropathy. *Transpl Infect Dis.* 2013;15(6):E270-E272. doi: 10.1111/tid.12142. 20. Oikawa M, Hatakeyama S, Fujita T, et al. BK virus-associated urothelial carcinoma of a ureter graft in a renal transplant recipient: a case report. *Transplant Proc.* 2014;46(2):616-619. doi: 10.1016/j.transproceed.2013.09.037.

21. Kenan DJ, Mieczkowski PA, Burger-Calderon R, Singh HK, Nickeleit V. The oncogenic potential of BK-polyomavirus is linked to viral integration into the human genome. *J Pathol.* 2015;237(3):379-389. doi: 10.1002/path.4584.

22. Alexiev BA, Drachenberg CB, Papadimitriou JC. Polyomavirus-cystitis associated with in situ and invasive urothelial carcinoma in a heart transplant recipient: evidence suggesting sequential progression/evolution from infection to carcinoma. *Transplantation*. 2015;99(1):e3-e4. doi: 10.1097/TP.000000000000509.

23. Tognon M, Corallini A, Martini F, Negrini M, Barbanti-Brodano G. Oncogenic transformation by BK virus and association with human tumors. *Oncogene*. 2003;22(33):5192-5200.

24. Dalianis T, Hirsch HH. BK polyomavirus and transformation. In: Robertson ES, editor. Cancer Associated Viruses. New York, NY: Springer Science & Business Media; 2012:419-432.

25. Weinreb DB, Desman GT, Amolat-Apiado MJ, Burstein DE, Godbold JH Jr, Johnson EM. Polyoma virus infection is a prominent risk factor for bladder carcinoma in immunocompetent individuals. *Diagn Cytopathol.* 2006;34(3):201-203.

26. Sanjuan NA. The paradox of polyomavirus. lytic infection or tumor induction? Medicina (B Aires). 2004;64(1):73-76. [article in Spanish]

27. Buch MH, Liaci AM, O'Hara SD, Garcea RL, Neu U, Stehle T. Structural and functional analysis of murine polyomavirus capsid proteins establish the determinants of ligand recognition and pathogenicity. *PLoS Pathog.* 2015;11(10):e1005104. doi: 10.1371/journal.ppat.1005104.