

# Second-Generation Brutons Tyrosine Kinase Inhibitors

Sarmen Sarkissian, MD, and Susan M. O'Brien, MD

## Abstract

Bruton tyrosine kinase (BTK) has been implicated in the pathogenesis of chronic lymphocytic leukemia (CLL) and other B-cell lymphoproliferative disorders. The first-generation BTK inhibitor, ibrutinib, has shown remarkable clinical activity in CLL, mantle cell lymphoma, marginal zone lymphoma, and Waldenström macroglobulinemia, both as monotherapy and as part of combination therapy. Despite the high response rates, subsequent resistance as well as adverse effects remain challenges. Numerous second-generation BTK inhibitors are in clinical trials at varying stages of development. In this review, we discuss 3 of these compounds: acalabrutinib (ACP-196), BGB-3111, and ONO/GS-4059.

*AJHO*. 2017;13(9):29-34

clinical responses in canine non-Hodgkin lymphoma (NHL) models. This was later followed by studies in human-derived chronic lymphocytic leukemia (CLL) cell lines, proving its attractiveness as a therapeutic target for B-cell malignancies.<sup>6-12</sup>

## Efficacy

In 2012, the clinical efficacy of ibrutinib was first reported in a phase I trial consisting of 50 patients with relapsed/refractory (r/r) B-cell malignancies; the objective response rate (ORR) was 60% (69% for CLL and for small lymphocytic leukemia [SLL]). A subsequent phase I/II trial in CLL/SLL resulted in an ORR of 71%. The superiority of ibrutinib over ofatumumab in the r/r setting was demonstrated in the RESONATE study, showing a 90% versus 81% overall survival at 12 months. The response rate was also significantly higher with ibrutinib, with an ORR of 42.6% versus 4.1%.<sup>13-15</sup>

Several combination studies have been published. A combination study of ibrutinib and ofatumumab in CLL, SLL, and prolymphocytic leukemia in 3 dosing cohorts showed response rates of 100%, 79%, and 71% for the ibrutinib lead-in, combination lead-in, and ofatumumab lead-in cohorts, respectively. A study of ibrutinib with rituximab in high-risk patients provided an ORR of 95%. A randomized trial comparing bendamustine/rituximab plus or minus ibrutinib showed improvements in median progression-free survival (PFS) (13.3 months vs not reached) and ORR (82.7% vs 67.8%).<sup>16-18</sup>

In the treatment-naïve population, the randomized phase III trial of ibrutinib versus chlorambucil showed a 13% improvement in 24-month survival and 51% improvement in ORR. Ibrutinib has efficacy in the high-risk *TP53* mutated/17p deleted population as well. A phase II study indicated 97% and 80% ORRs in treatment-naïve and r/r patients, respectively. In the largest trial ever reporting outcomes in r/r patients with CLL and 17p deletion, 134 patients treated with ibrutinib had an 83% ORR and a 24-month PFS of 63%. The tolerability of ibrutinib as frontline therapy for CLL in an older population was also studied, revealing a very manageable adverse event (AE) profile.<sup>19-22</sup>

The data in other B-cell malignancies are also impressive. In relapsed mantle cell lymphoma (MCL), the first phase II trial with ibrutinib provided an ORR of 68%; in combination with

## Introduction

First described in the 1950s, the syndrome Bruton X-linked agammaglobulinemia was identified as a humoral immunodeficiency. The Bruton tyrosine kinase (BTK) gene, *BTK*, was cloned in 1993. It consists of 19 exons, and more than 250 mutations have been described in the BTKBase database to date. The *BTK* gene is on the q arm of the X chromosome at the g21.1-22 locus, and the BTK protein is the transcription product. BTK is a member of the TEC family of tyrosine kinases and is found in the cytoplasm. The protein is normally phosphorylated after the B-cell receptor is activated, and it is involved in the signaling pathway that leads to B-cell differentiation (Figure). A deficiency of a functional protein leads to an arrest in B-cell maturation.<sup>1,5</sup>

After the gene was cloned, numerous studies further defined its role in B-cell receptor (BCR) signaling and maturation. It promotes cell cycle progression and blunts apoptosis via the *Nf- $\kappa$ B* and *BCL-X1* pathways, respectively. The dependence of mature B cells on BTK signaling was shown in mouse models. In vitro studies revealed not only the necessity of the BCR for cell survival, but a viable small molecule target. This led to the first published report of the irreversible BCR inhibitor, ibrutinib, which selectively blocked B-cell receptor signaling and induced

**TABLE. Summaries of Highlighted Trials Related to BTK Inhibitors (other than ibrutinib)**

Compound	Phase of Trial	N	Tumor Subtypes	Line	Overall Response Rate	Grade 3/4 Toxicities
ONO/GS-4059	I	90	CLL/SLL, MCL, DLBCL, FL, MZL, and WM	r/r	CLL: 96% (n = 24/25); MCL: 92% (n = 11/12); DLBCL: 35% (n = 11/31)	n = 42
BGB-3111	I	25	CLL/SLL, MCL, DLBCL, HCL, FL, MZL, and WM	r/r	Entire group: 88% (n = 22/25)	n = 21
Acalabrutinib	I/II	61	CLL/SLL	r/r	95% (n = 53/61)	n = 11

CLL indicates chronic lymphocytic leukemia; DLBCL, diffuse large B cell lymphoma; FL, follicular lymphoma; MCL, mantle cell lymphoma; MZL, marginal zone lymphoma; r/r, relapsed/refractory; SLL, small lymphocytic leukemia; WM, Waldenström macroglobulinemia.

rituximab, the ORR was 88%. In another randomized trial in r/r MCL with temsirolimus as the comparator arm, the PFS improved by 8 months—14.6 versus 6.2 months—and the ORR improved from 40% to 72%.<sup>23,24</sup>

In r/r Waldenström macroglobulinemia (WM), ibrutinib also has activity. The expanded phase II study for r/r WM showed an ORR of 90.5%. Patients with the mutation MYD88 L265P and CXCR WT had an ORR of 100%. Patients with the mutations MYD88 L265P and CXCR4 WHIM (indicating warts, hypogammaglobulinemia, infections, and myelokathexis) had an ORR of 85.7%, and those with MYD88 WT CXCR4 WHIM had an ORR of 71.4%. It has also shown early promise in r/r diffuse large B-cell lymphoma (DLBCL). When combined with rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone (R-CHOP), the ORR was 100% in the activated B-cell (ABC) subtype. As a single agent, the ORR in the ABC subtype of r/r DLBCL was 25%.<sup>25-27</sup>

### Resistance

#### Chronic Lymphocytic Leukemia

Primary and secondary resistance to therapy remains a clinical challenge. Whole genome sequencing performed in 5 of 6 longitudinally-followed patients with CLL who developed ibrutinib resistance showed a C481S substitution in *BTK*. In another analysis, 10 of 10 patients with CLL and Richter transformation had either a *BTK* C481S mutation or a *PLCG2* mutation. The presence of the *BTK* C481S mutation leads to loss of irreversible binding. The C481S mutation has 25-fold less sensitivity to ibrutinib than does the wild type.

Mutated *BTK* activation causes downstream gene expression, cell signaling, and cell behavior changes promoting cell survival. These mutations retain sustained sensitivity to numerous agents, including dasatinib, idelalisib, and SYK inhibitors.<sup>28-32</sup>

#### Mantle Cell Lymphoma

Primary resistance in MCL appears to be more complicated. Whole exome/transcriptome profiling of serial biopsies, from ibrutinib initiation until drug resistance, in a patient with MCL

showed the evolution of the *BTK* C481S mutation. Data revealed amplified activation of the *BTK*, *AKT*, and *CXCR4-CXCL12* pathways, thought to be mediators of secondary resistance. Other experiments have shown that in ibrutinib-resistant MCL cell lines, cell proliferation is mediated through a nonclassical NF-κB pathway, via *TRAF2* and *TRAF3* mutations.<sup>33,34</sup>

#### Diffuse Large B-cell Lymphoma

DLBCL comprises a genetically heterogeneous group of B-cell lymphomas. The activated B cell–like (ABC) subtype has a gain-of-function *CD79A/B* ITAM motif mutation. MYD88 L265P mutations are also seen more commonly in the ABC than the germinal center subtype. The response rates to ibrutinib for ABC and germinal center B cell–like (GCB) tumors were 38% and 5%, respectively. Tumors with a mutation in MYD88 or *CD79A/B* WT were noted to be more sensitive to ibrutinib than tumors without such mutations.<sup>35-37</sup>

#### Waldenström Macroglobulinemia

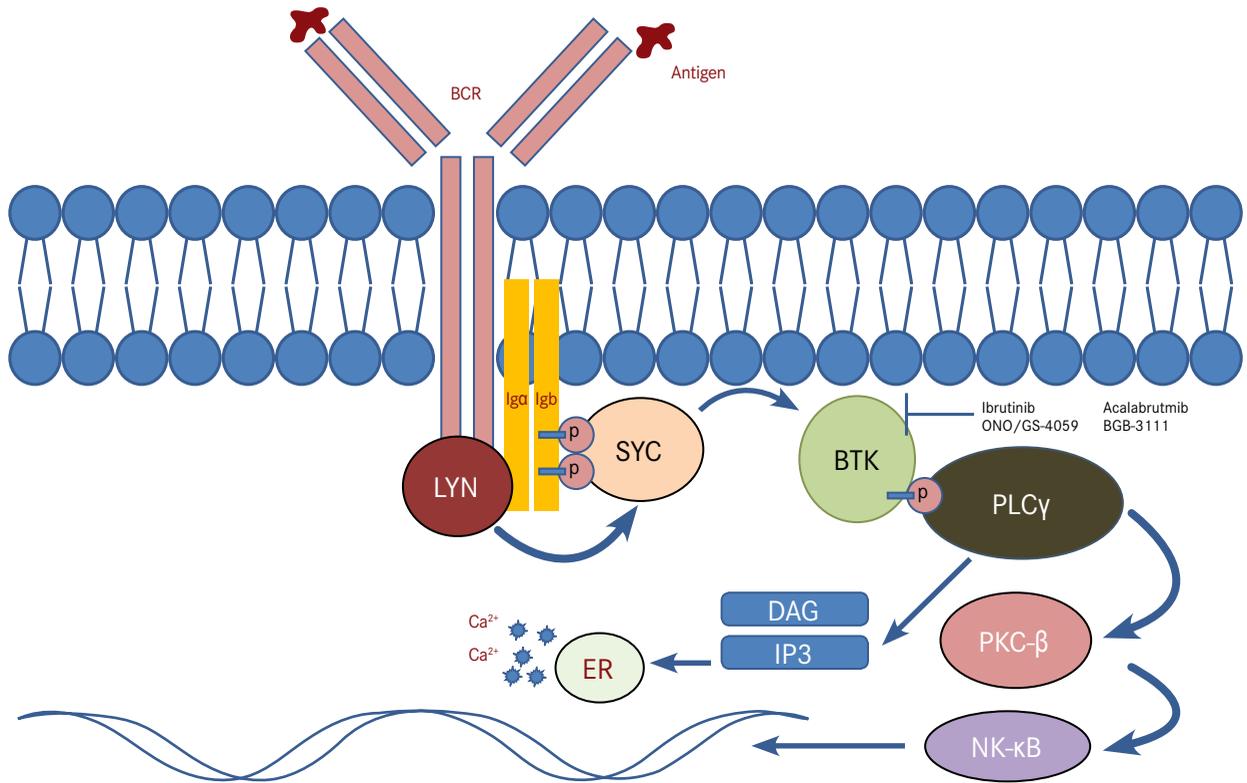
In the phase II study of ibrutinib in r/r WM, the patients with the mutations MYD88 L265P and CXCR4 WHIM had an ORR of 85.7%, and those with MYD88 WT CXCR4 WHIM had a lower ORR of 71.4%. It is thought the WHIM-like CXCR4 S338X mutation (CXCR4 WHIM) allows for a bypass circuit to the mTOR-AKT pathway in the setting of the MYD88 L265P mutation, thus conferring resistance to ibrutinib.<sup>38,39</sup>

#### Other BTK Inhibitors

Highlighted trials related to BTK inhibitors, other than ibrutinib, are highlighted in the **Table**.

#### Acalabrutinib

Acalabrutinib (ACP-196) is a second-generation BTK inhibitor that irreversibly binds the BTK cysteine domain. It has higher selectivity for BTK as compared with other kinases, including MEC, PDGFR, EGFR, ITK, and others. The selectivity is thought to be due to acalabrutinib’s propiolamide side group versus ibrutinib’s acrylamide side group. The propiolamide

**FIGURE. BTK Signaling Pathway in the B Lymphocyte**

Activation of Bruton tyrosine kinase (BTK) by the antigen-bound activated B-cell receptor leads to phosphorylation of BTK. This in turn promotes both cell survival and antiapoptotic mechanisms. Inhibition of this protein then causes modulation of this signaling pathway, leading to clinical responses seen in several B-cell malignancies.

side group may allow fewer off-target interactions with other kinases such as ITK, EGFR, ERBB2/4, JAK3, LYN, and SRK. While the half maximal inhibitory concentration ( $IC_{50}$ ) for BTK is roughly 3 times that of ibrutinib, the  $IC_{50}$  against other kinases is much higher.<sup>40-42</sup>

In a phase I/II report of 61 patients with r/r CLL, the ORR at a median follow-up of 14.3 months was 95%, with 53 of 61 patients still on therapy. The phase I portion used 100 mg, 175 mg, 250 mg, and 400 mg once-daily dosing, while the phase II portion used 100 mg twice-daily dosing. The response rate in the 17p-deleted patient cohort was 100%. The response rate in those who had received idelalisib was 100%. Grade 3/4 AEs included diarrhea (n = 1; 2%), weight gain (n = 1; 2%), arthralgia (n = 1; 2%), pyrexia (n = 2; 3%), fatigue (n = 2; 3%), and hypertension (n = 4; 7%). No Richter transformation occurred and only 1 patient progressed.<sup>43</sup>

#### ONO/GS-4059

ONO/GS-4059 is a covalent inhibitor with a greater selec-

tivity for BTK than for LCK, FYN, LYN, and ITK. In vitro studies showed that the 50% inhibitory concentration of BTK by ONO/GS-4059 was only 2 nmol/L. ONO/GS-4059 showed promising preclinical efficacy, and this prompted the development of a phase I study in r/r B-cell malignancies.<sup>44-46</sup>

In a phase I study of ONO/GS-4059, 90 patients with a variety of B-cell malignancies, including CLL/SLL, MCL, DLBCL, follicular lymphoma (FL), marginal zone lymphoma (MZL), and WM, were enrolled in a 3+3 dose-escalation study. Doses ranged from 20 mg to 600 mg daily. There were 2 parallel dose-escalation groups, 1 for NHL and 1 for CLL. The overall estimated mean PFS rates in CLL, MCL, and DLBCL were 874, 341, and 54 days, respectively. There was a 96% ORR in patients with CLL (24/25 patients). After 29 months, 34 of 90 patients remained on study, including 21 patients with CLL as well as 13 patients with NHL. Of the patients with CLL, 13 had loss of *TP53* and 21 had unmutated IGHV gene segments. All 12 patients with *TP53*/17p deletion or *TP53* mutations had a response, with 9 remaining on study. In the MCL

group, 11 of 12 evaluable patients showed a response (n = 6, partial response [PR]; and n = 5, complete response/unconfirmed complete response [CR/CRu]). Eight of 12 patients remained on study. In the DLBCL cohort (31/35 ABC), 11 of 31 (35%) showed a response (9 PRs and 2 CR/CRus).<sup>47</sup>

A total of 171 AEs were reported, 75% of which were grade 1/2. Grade 3/4 toxicities included neutropenia (n = 9; 10%), anemia (n = 12; 13.3%), and thrombocytopenia (n = 12; 13.3%). Ten patients discontinued therapy because of AEs/dose-limiting toxicities (DLTs) (n = 5 [CLL] and n = 5 [other NHL]). There was 1 reported case of new-onset atrial fibrillation, which was not thought to be drug related. There were no grade 3/4 cases of diarrhea or arthralgia. In the CLL dose-expansion cohort, the maximum tolerated dose (MTD) of 600 mg daily was reached without DLTs. In the lymphoma cohort, an MTD of 480 mg once daily was reached because 4 patients experienced DLTs (n = 1 each, for urticarial reaction, maculopapular cutaneous rash, nonimmune drug reaction, and cutaneous rash). It is unclear why the DLTs were seen in NHL versus CLL. Other grade 3/4 toxicities included 5 lower respiratory tract infections as well as 1 urticarial rash.

### BGB-3111

BGB-3111 is an irreversible inhibitor of BTK. In preclinical models of MCL and DLBCL, it inhibited BTK auto-phosphorylation and cell proliferation. Cell culture and biochemical assays with BGB-3111 showed less off-target kinase inhibition than was seen with ibrutinib. The IC<sub>50</sub> ratio of BGB-3111 to ibrutinib for non-BTK kinases (TEK, EGFR, ITK, JAK3, and HER2) ranged from 2- to 70-fold. The IC<sub>50</sub> ratio:ratio BGB-3111:ibrutinib for BTK was 0.5. BGB-3111 was nearly 10 times weaker at inhibiting rituximab-induced antibody-dependent cell-mediated cytotoxicity. It has 3-fold higher potency compared with ibrutinib; it also demonstrated dose-dependent antitumor effects against REC-1 MCL xenografts, and prolongation of overall survival in a xenograft model.<sup>48</sup>

In the phase I trial, 25 patients with various B-cell malignancies were enrolled. Diagnoses included CLL (n = 8), MCL (n = 6), WM (n = 6), DLBCL (n = 2), FL (n = 1), MZL (n = 1), and HCL (n = 1). Patients were treated in 5 dosing cohorts: 40, 80, 160, or 320 mg once daily, or 160 mg twice daily. Pharmacokinetic studies showed a sustained 24-hour BTK occupancy in peripheral blood mononuclear cells as well as in lymph nodes for all patients at all doses. Twenty-two of 25 patients had a response, with 15 PRs, 1 CR, and 6 stable disease. There were 21 grade 3/4 AEs, 3 of which were believed to be drug-related. There were no cases of atrial fibrillation/flutter.<sup>49</sup>

### Conclusions

The BTK pathway plays a crucial role in the pathogenesis of B-cell malignancies. Targeting BTK has changed the treatment landscape for B-cell malignancies. Although BTK inhibition has found

a role in CLL, WM, MZL, and MCL, its application to other B-cell malignancies is under evaluation.

Better understanding of the structural characteristics of ibrutinib has allowed for the development of other potent, selective inhibitors. As the data on long-term efficacy and safety of ibrutinib mature, resistance and AEs remain as clinical challenges. Further elucidating resistance mechanisms will guide the development of the next generation of BTK inhibitors. Randomized trial data are needed to better address their true efficacy and AE profile. Lastly, their role as monotherapy and in combination therapy will help fine-tune the treatment armamentarium for B-cell malignancies.

**Affiliations:** Both Sarmen Sarkissian, MD, and Susan M. O'Brien, MD, are with the Department of Hematology and Oncology, University of California at Irvine. Dr O'Brien is also with UC Irvine's Chao Family Comprehensive Cancer Center.

**Corresponding author:** Susan M. O'Brien, MD, Chao Family Comprehensive Cancer Center, 101 The City Dr, Orange, CA 92868. Tel: (714) 456-7511; E-mail: Obrien@uci.edu.

**Financial disclosures:** None.

### References

1. Parslow TG, Stites DP, Terr AI, Imboden JB. *Medical Immunology*. 10th ed. New York, NY: McGraw-Hill; 2001.
2. Virella G, ed. *Medical Immunology*. 6th ed. Boca Raton, FL: CRC Press; 2007.
3. Paul WE. *Fundamental Immunology*. 5th ed. Philadelphia, PA: Lippincott Williams & Wilkins; 2003.
4. Rich R. *Clinical Immunology Principles and Practices*. 2nd ed. London, UK: Mosby; 2001.
5. Hyvönen M, Saraste M. Structure of the PH domain and Btk motif from Bruton's tyrosine kinase: molecular explanations for X-linked agammaglobulinemia. *EMBO J*. 1997;16(12):3396-3404.
6. Satterthwaite AB, Witte ON. The role of Bruton's tyrosine kinase in B-cell development and function: a genetic perspective. *Immunol Rev*. 2000;175:120-127.
7. Khan WN. Regulation of B lymphocyte development and activation by Bruton's tyrosine kinase. *Immunol Res*. 2001;23(2-3):147-156.
8. Humphries LA, Dangelmaier C, Sommer K, et al. Tec kinases mediate sustained calcium influx via site-specific tyrosine phosphorylation of the phospholipase Cgamma Src homology 2-Src homology 3 linker. *J Biol Chem*. 2004;279(36):37651-37661.
9. Satterthwaite AB, Cheroutre H, Khan WN, et al. Btk dosage determines sensitivity to B cell antigen receptor cross-linking. *Proc Natl Acad Sci U S A*. 1997;94(24):13152-13157.
10. Davis RE, Ngo VN, Lenz G, et al. Chronic active B-cell-

receptor signaling in diffuse large B-cell lymphoma. *Nature*. 2010;463(7277):88-92. doi: 10.1038/nature08638.

11. Honigberg LA, Smith AM, Sirisawad M, et al. The Bruton tyrosine kinase inhibitor PCI-32765 blocks B-cell activation and is efficacious in models of autoimmune disease and B-cell malignancy. *Proc Natl Acad Sci U S A*. 2010;107(29):13075-13080. doi: 10.1073/pnas.1004594107.
12. Herman SE, Gordon AL, Hertlein E, et al. Bruton tyrosine kinase represents a promising therapeutic target for treatment of chronic lymphocytic leukemia and is effectively targeted by PCI-32765. *Blood*. 2011;117(23):6287-6296. doi: 10.1182/blood-2011-01-328484.
13. Advani RH, Buggy JJ, Sharman JP, et al. Bruton tyrosine kinase inhibitor ibrutinib (PCI-32765) has significant activity in patients with relapsed/refractory B-cell malignancies. *J Clin Oncol*. 2013;31(1):88-94. doi: 10.1200/JCO.2012.42.7906.
14. Byrd JC, Furman RR, Coutre SE, et al. Targeting BTK with ibrutinib in relapsed chronic lymphocytic leukemia. *N Engl J Med*. 2013;369(1):32-42. doi: 10.1056/NEJMoa1215637.
15. Byrd JC, Brown JR, O'Brien S, et al; RESONATE Investigators. Ibrutinib versus ofatumumab in previously treated chronic lymphoid leukemia. *N Engl J Med*. 2014;371(3):213-223. doi: 10.1056/NEJMoa1400376.
16. Jaglowski SM, Jones JA, Nagar V, et al. Safety and activity of BTK inhibitor ibrutinib combined with ofatumumab in chronic lymphocytic leukemia: a phase 1b/2 study. *Blood*. 2015;126(7):842-850. doi: 10.1182/blood-2014-12-617522.
17. Burger JA, Keating MJ, Wierda WG, et al. Safety and activity of ibrutinib plus rituximab for patients with high-risk chronic lymphocytic leukaemia: a single-arm, phase 2 study. *Lancet Oncol*. 2014;15(10):1090-1099. doi: 10.1016/S1470-2045(14)70335-3.
18. Chanan-Khan A, Cramer P, Demirkan F, et al; HELIOS investigators. Ibrutinib combined with bendamustine and rituximab compared with placebo, bendamustine, and rituximab for previously treated chronic lymphocytic leukaemia or small lymphocytic lymphoma (HELIOS): a randomised, double-blind, phase 3 study. *Lancet Oncol*. 2016;17(2):200-211. doi: 10.1016/S1470-2045(15)00465-9.
19. Burger JA, Tedeschi A, Barr PM, et al; RESONATE-2 Investigators. Ibrutinib as initial therapy for patients with chronic lymphocytic leukemia. *N Engl J Med*. 2015;373(25):2425-2437. doi: 10.1056/NEJMoa1509388.
20. Farooqui MZ, Valdez J, Martyr S, et al. Ibrutinib for previously untreated and relapsed or refractory chronic lymphocytic leukaemia with TP53 aberrations: a phase 2, single-arm trial. *Lancet Oncol*. 2015;16(2):169-176. doi: 10.1016/S1470-2045(14)71182-9.
21. O'Brien S, Jones JA, Coutre SE, et al. Ibrutinib for patients with relapsed or refractory chronic lymphocytic leukaemia with 17p deletion (RESONATE-17): a phase 2, open-label, multicentre study. *Lancet Oncol*. 2016;17(10):1409-1418. doi: 10.1016/S1470-2045(16)30212-1.
22. O'Brien S, Furman RR, Coutre SE, et al. Ibrutinib as initial therapy for elderly patients with chronic lymphocytic leukaemia or small lymphocytic lymphoma: an open-label, multicentre, phase 1b/2 trial. *Lancet Oncol*. 2014;15(1):48-58. doi: 10.1016/S1470-2045(13)70513-8.
23. Wang ML, Rule S, Martin P, et al. Targeting BTK with ibrutinib in relapsed or refractory mantle-cell lymphoma. *N Engl J Med*. 2013;369(6):507-516. doi: 10.1056/NEJMoa1306220.
24. Dreyling M, Jurczak W, Jerkeman M, et al. Ibrutinib versus temsirolimus in patients with relapsed or refractory mantle-cell lymphoma: an international, randomised, open-label, phase 3 study. *Lancet*. 2016;387(10020):770-778. doi: 10.1016/S0140-6736(15)00667-4.
25. Treon SP, Tripsas CK, Meid K, et al. Ibrutinib in previously treated Waldenström's macroglobulinemia. *N Engl J Med*. 2015;372(15):1430-1440. doi: 10.1056/NEJMoa1501548.
26. Younes A, Thieblemont C, Morschhauser F, et al. Combination of ibrutinib with rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone (R-CHOP) for treatment-naïve patients with CD20-positive B-cell non-Hodgkin lymphoma: a non-randomised, phase 1b study. *Lancet Oncol*. 2014;15(9):1019-1026. doi: 10.1016/S1470-2045(14)70311-0.
27. Wilson WH, Young RM, Schmitz R, et al. Targeting B cell receptor signaling with ibrutinib in diffuse large B cell lymphoma. *Nat Med*. 2015;21(8):922-926. doi: 10.1038/nm.3884.
28. Woyach JA, Furman RR, Liu TM, et al. Resistance mechanisms for the Bruton's tyrosine kinase inhibitor ibrutinib. *N Engl J Med*. 2014;370(24):2286-2294. doi: 10.1056/NEJMoa1400029.
29. Byrd JC. Richter's transformation and CLL after ibrutinib: BTK and PLCg2 mutations. In: American Society of Hematology (ASH), ed. *American Society of Hematology: Meeting on Lymphoma Biology*. Colorado Springs, CO: ASH; 2014:14.
30. Furman RR, Cheng S, Lu P, et al. Ibrutinib resistance in chronic lymphocytic leukemia. *N Engl J Med*. 2014;370(24):2352-2354. doi: 10.1056/NEJMoa1402716.
31. Cheng S, Guo A, Lu P, et al. Functional characterization of BTK(C481S) mutation that confers ibrutinib resistance: exploration of alternative kinase inhibitors. *Leukemia*. 2014;28(3):649-657. doi: 10.1038/leu.2013.358.
32. Liu Q, Sabnis Y, Zhao Z, et al. Developing irreversible inhibitors of the protein kinase cys200. *Chem Biol*. 2013;20(2):146-159. doi: 10.1016/j.chembiol.2012.12.006.
33. Chiron D, Di Liberto M, Martin P, et al. Cell-cycle reprogramming for PI3K inhibition overrides a relapse-specific C481S BTK mutation revealed by longitudinal functional genomics in mantle cell lymphoma. *Cancer Discov*. 2014;4(9):1022-1035. doi: 10.1158/2159-8290.CD-14-0098.
34. Rahal R, Frick M, Romero R, et al. Pharmacological and genomic profiling identifies Nf- $\kappa$ B-targeted treatment strategies for mantle cell lymphoma. *Nat Med*. 2014;20(1):87-92. doi: 10.1038/nm.3435.

35. Davis RE, Ngo VN, Lenz G, et al Chronic active B-cell-receptor signaling in diffuse large B-cell lymphoma. *Nature*. 2010;463(7277):88-92. doi: 10.1038/nature08638.
36. Staudt LM. Oncogenic activation of NFkappaB. *Cold Spring Harb Perspect Biol*. 2010;2(6):a000109. doi: 10.1101/cshperspect.a000109.
37. Wilson WH, Gerecitano JF, Goy A, et al. The Bruton's tyrosine kinase (BTK) inhibitor, ibrutinib (PCI-32765), has preferential activity in the ABC subtype of relapsed/refractory de novo diffuse large B-cell lymphoma (DLBCL): interim results of a multicenter, open-label, phase 2 study [abstract]. Abstract presented at: 54<sup>th</sup> ASH Annual Meeting and Exposition; December 8-11, 2012; Atlanta, GA. Abstract 686.
38. Treon SP, Tripasas CK, Yang G, et al. A prospective multicenter study of the Bruton's tyrosine kinase inhibitor ibrutinib in patients with relapsed or refractory Waldenström's macroglobulinemia [abstract]. Abstract presented at: 55<sup>th</sup> Annual Meeting and Exposition; December 7-10, 2013; New Orleans, LA. Abstract 251.
39. Cao Y, Hunter ZR, Liu X, et al. The WHIM-like CXCR4(S338X) somatic mutation activates AKT and ERK, and promotes resistance to ibrutinib and other agents used in the treatment of Waldenström's macroglobulinemia. *Leukemia*. 2015;29(1):169-76. doi: 10.1038/leu.2014.187.
40. Maly J, Blachly JS. Chronic lymphocytic leukemia: exploiting vulnerabilities with targeted agents. *Curr Hematol Malig Rep*. 2016;11(1):52-60. doi: 10.1007/s11899-016-0299-0.
41. Covey T, Barf T, Gulrajani et al. ACP-196: a novel covalent Bruton's tyrosine kinase (Btk) inhibitor with improved selectivity and in vivo target coverage in chronic lymphocytic leukemia (CLL) patients. *Cancer Res*. 2015;75(Supplement 15):2596. doi: 10.1158/1538-7445.AM2015-2596.
42. Byrd JC, Wierda W, Jones J, et al. The Bruton tyrosine kinase (Btk) inhibitor ACP-196: marked activity in relapsed/refractory CLL with a favorable safety profile. *Blood*. 2015;126:831.
43. Byrd JC, Harrington B, O'Brien S, et al. Acalabrutinib (ACP-196) in relapsed chronic lymphocytic leukemia. *N Engl J Med*. 2016;374(4):323-332. doi: 10.1056/NEJMoa1509981.
44. Yasuhiro T, Yoshizawa T, Daub H, et al. ONO-WG-307, a novel, potent and selective inhibitor of Bruton's tyrosine kinase (Btk), results in sustained inhibition of the ERK, AKT and PKD signaling pathways. In: Proceedings of the American Association for Cancer Research 103<sup>rd</sup> Annual Meeting; March 31-April 4, 2012; Chicago, IL. Abstract 2021.
45. Kozaki R, Hutchinson C, Sandrine J, Dyer MJS. Kinome reprogramming in DLBCL by the BTK-specific inhibitor ONO-4059 highlights synergistic combinations for clinical application [abstract]. *Haematologica*. 2014;99(S1):137-138. Abstract P431.
46. Kozaki R, Yoshizawa T, Tohda S, et al. Development of a Bruton's tyrosine kinase (Btk) inhibitor, ONO-WG-307: efficacy in ABC-DLBCL xenograft model—potential treatment for B-cell malignancies [abstract]. *Blood*. 2011;118(21):Abstract 3731.
47. Walter HS, Rule SA, Dyer MJ, et al. A phase 1 clinical trial of the selective BTK inhibitor ONO/GS-4059 in relapsed and refractory mature B-cell malignancies. *Blood*. 2016;127(4):411-419.
48. Na L, Zhijian S, Ye L, et al. BGB-3111 is a novel and highly selective Bruton's tyrosine kinase (BTK) inhibitor. In: Proceedings of the American Association for Cancer Research 106<sup>th</sup> Annual Meeting; April 18-22, 201; Philadelphia, PA. Abstract 2597.
49. Tam C, Grigg AP, Opat S, et al. The BTK inhibitor, Bgb-3111, is safe, tolerable, and highly active in patients with relapsed/refractory B-cell malignancies: initial report of a phase 1 first-in-human trial. *Blood*. 2015;126(23):832.